



## FERROPTOSIS AND ITS POTENTIAL ROLE IN CARDIOVASCULAR DISEASES

Yavor Kashlov, Maria Dimova

*Department of Internal Medicine, UMHAT St. Marina, Faculty of Medicine, Medical University, Varna, Bulgaria.*

### ABSTRACT

Iron (Fe) is the fourth most common element in the earth's crust, and plays a significant role in the human body. It is important for cell survival due to its involvement in the transport of oxygen, in DNA biosynthesis, adenosine triphosphate (ATP) synthesis, as well as an auxiliary factor of various proteins in the tricarboxylic acid cycle (TCA). It has also been found that iron is closely related to the onset and progression of various cancers and disturbances in its metabolism can stimulate the growth of tumor cells. In addition, divalent iron significantly accelerates the lipid peroxidation of saturated fatty acids.

**Keywords:** Ferroptosis, adenosine triphosphate (ATP), erastin, cell death, apoptosis, necrosis,

### INTRODUCTION

During the oxidative phosphorylation in mitochondria, which is an iron-dependent process, along with the production of ATP, reactive oxygen species (ROS) are also generated. The generated in large quantities oxygen radicals, exceeding the capacity of the reductase mechanisms of the cells, can result in direct or indirect injury to different molecules such as proteins, nucleic acids and lipids [1], and consequently also to cell death. This newly discovered form of cell death is called ferroptosis. It differs from apoptosis and necrosis and is a result of the accumulation of iron-dependent lipid peroxidase [2].

Typical morphological changes are shrinking of cell volume and increase in the density of the mitochondrial membrane [2]. Ferroptotic cell death is induced by two different classes of small molecules. Class I inducers of ferroptosis include erastin, sulfasalazine (SAS), dihydroxyphenyl-imino-2-imidazolidine (DPI2) and buthionine sulfoximine, which reduce the intracellular glutathione level, causing an imbalance between oxidation-reduction in the cells. Class 2 ferroptosis inducers include Ras selective lethal compound (RSL3), DPI7, DPI10, DPI12, DPI13, etc. that directly inhibit glutathione peroxidase 4 (GPx4) [3] and eventually lead to accumulation of lipid peroxides in the cell. In addition, ferroptosis can be induced by various drugs (e.g., sorafenib,

artemisinin and its derivatives) [4, 5].

Regulators of ferroptosis are mainly the mevalonate pathway, the sulfur transfer signaling pathways and the heat shock protein 1- heat shock factor 1 (HSF1-HSPB1) system [6]. This type of cell death can occur during various physiological and pathological processes in humans and animals. The research describes the involvement of ferroptosis in various disorders in humans. In particular, by regulating the growth and proliferation of tumor cells, ferroptosis plays an important role in the development and progression of various cancers. Up until now susceptibility of tumor cells to ferroptosis has been reported for pancreatic ductal carcinoma, renal cell carcinoma (RCC) and hepatocellular carcinoma (HCC) [3, 7, 8].

### Discovery of ferroptosis

Historically, the inducers of ferroptosis were found initially, before the process was described. In a 2003 study using large-scale screening to investigate the cytolytic effect of various chemical compounds on tumor cells, Stockwell et al. identified a new chemical compound, called erastin. This compound has the ability to induce death in tumor cells with a RAS mutation, in a manner different from apoptosis [9]. In 2008, Stockwell et al. discovered two new molecules - RSL3 and RSL5, which have the same effect as erastin. In this study, it was found that the observed cell death can be inhibited by the iron chelator - desferrioxamine B-methanesulfonate (DFOM) and the antioxidant vitamin E [10], thus confirming that this form of cell death is associated with the intracellular ferrous ion and the reactive oxygen species. In 2012, Stockwell et al. used the term "ferroptosis" to describe this type of cell death, caused by the accumulation of iron-dependent lipid peroxides [2]. Later, other compounds have been determined as well (sorafenib [11], artemisinins [5,7], cyclic peroxide 1,2-dioxolane - FINO2) [12], that have the ability to induce ferroptosis. In addition, the activation mechanisms for ferroptosis molecular systems Xc- (cystine-glutamate antiporter that is chloride dependent) and GPx4 [3,13] have been discovered. Recently, a relatively detailed summary of ferroptosis inducers, inhibitors and regulatory molecules has been published [14].

### **Differences between ferroptosis and apoptosis / necrosis**

Cell death, which represents the end-stage of the cell cycle, is caused by various cytotoxic exogenous or endogenous substances. In 1972, Kerr et al. described “automatically programmed” cell death, which they called apoptosis [15].

Later, by using electron microscopy, Schweichel and Merker observed types of cell death of embryonic cells in rats that were treated or not with lethal to the embryo substances. They divided the cell death into three types [16].

Clark named the third type – necrosis [17], which is a passive form of cell death. With the advance of scientific research in this area, various types of cell death, such as pyroptosis, necroptosis, parthanatos, autophagy, oncosis and ferroptosis were discovered. The latter form differs significantly from other types such as apoptosis, necrosis and autophagy including by its morphological, biochemical and genetic features [2,10].

Ferroptosis does not lead to morphological changes, such as the condensation of the chromatin that occurs during apoptosis, loss of the plasma membrane integrity of the cell that occurs during necrosis or the formation of autophagic vacuoles that appear during autophagy. Instead, ferroptosis is characterized primarily by a reduction in the volume of mitochondria and an increase in density with subsequent rupture of the external mitochondrial membrane [2].

The induction of ferroptosis is done by different types of low molecular weight molecules such as erastin and RSL3. It is interesting to note that it cannot be induced by substances that are triggers for apoptosis and necrosis<sup>5</sup>, suggesting that the mechanism underlying ferroptosis differs from that of apoptosis and necrosis. In addition, the inducers of ferroptosis generally differ from those of other newly discovered forms of cell death, such as necroptosis and pyroptosis [18].

A recent study has found a link between ferroptosis and autophagy, which share some common mechanisms [19]. Therefore, the relationship between ferroptosis and other types of cell death remains to be elucidated.

The cysteine / glutamate transport system Xc- is a membrane Na<sup>+</sup>-dependent cysteine-glutamate exchange transporter, which is a disulfide-linked heterodimer consisting of a light (xCT, SLC7A11) and heavy chain (CD98hc, SLC3A2) [20]. It transports extracellular cysteine inside the cell [21], which is then transformed into cysteine used for glutathione synthesis (GSH). This molecule is critical for the protection of the cell against injury caused by oxidative stress [22].

In the central nervous system (CNS), the neurotoxicity of glutamate is oxidative iron-dependent [23, 24].

It can be inhibited by iron chelators and ferr-1 [2], which means that an involvement in ferroptosis is possible [23]. Previous studies have revealed that glutamate toxicity is due to an increased calcium influx in the cell or is a result of inhibition of the Xc- system by competitive inhibitors [21, 25]. However, Wolpaw et al. reported that calcium chelators did not affect ferroptosis [26]. The latter conclusion supports the thesis that the activation of glutamate receptors is not involved in ferroptosis [18].

### **Ferroptosis and cardiovascular system**

Despite the role that ferroptosis plays in the pathogenesis of various oncological diseases is being studied extensively, the role of ferroptosis in cardiovascular disease is not so well researched.

Initially, ferroptosis has been observed in cancer cells expressing an oncogenic Ras gene, and subsequently in other disorders such as Huntington’s disease and renal failure [27], but there are few studies on the role of the process in cardiovascular disease. Nevertheless, the study of iron homeostasis and the involvement of iron in the myocardial damage has a long history. The term “iron overload cardiomyopathy” has been introduced to designate a type of myocardial damage, resulting from the accumulation of iron in the myocardium, primarily due to genetically determined disorders of iron metabolism or as a result of multiple blood transfusions [28].

The clinical use of doxorubicin is limited due to its cardiotoxicity. In a study, the concomitant use of iron chelators together with doxorubicin resulted in limiting the cardiotoxicity, which raises the question of the likely involvement of iron in myocardial injury, mediated by the chemotherapeutic agent [29]. Nitenberg et al. showed that in patients with diabetes mellitus and heart failure (HF), the iron homeostasis is possibly impaired, and chelation therapy may improve coronary microvasculature [30].

In a recent study by Lapenna et al. [31] have been reported higher levels of low molecular weight iron (LMWI) - a redox-active catalytic form of iron, as well as higher levels of lipid and protein oxidation in the hearts of adult rabbits as compared to the younger controls. These results show that ferroptosis, characterized by an increased oxidative stress, is probably one of the major mechanisms of cell death leading to cardiac dysfunction. However, the iron homeostasis and associated with it oxidative stress are still not enough studied in heart failure caused by increased preload. Therefore, future research into potentially effective therapeutic strategies, aimed at influencing ferroptosis, is needed.

As a tissue that consumes a large amount of oxygen, the heart is rich in unsaturated fatty acids, which provides the pathophysiological basis for the generation of reactive oxygen species with subsequent myocardial damage [32].

In addition, in situations of stress, mitochondrial or functional impairment, the abundance of non-heme-bound iron in the myocardium can cause lipid peroxidation of the cell membrane through the Fenton reaction (reaction between hydrogen peroxide and ferrous ions) [32].

There are a number of studies that demonstrate the key role of oxidative stress in cardiomyocyte death. In addition, in some cardiovascular diseases such as myocardial infarction, in physiological processes such as aging has been described regional accumulation of iron in the myocardium [28-31]. Interestingly, iron chelators and antioxidant agents can effectively reduce the death of cardiomyocytes [28-31], suggesting that ferroptosis is a possible mechanism for cell death in cardiovascular diseases.

In animal models of myocardial ischemia-reperfusion injury, the blocking of the key molecule for ferroptosis - glutathione results in a reduction in myocardial injury [33]. In another interesting study, the role of ferroptosis in the induction of ischemia-reperfusion injury has been reported [34]. Initially, the iron load and the fibrosis were assessed in cardiomyocyte populations in mice, subjected to an ischemia-reperfusion injury. It is significant to mention the accumulation of iron in cardiomyocytes and the development of excessive fibrosis around the deposits, one week after ischemia-reperfusion injury (I / R injury). In order to assess the degree and presence of ferroptosis in these cardiomyocytes, the authors use separately three molecules (erastin, RSL3, and Fe<sup>3+</sup>), leading either to an increase in cellular iron levels or to increased iron-mediated oxidative stress. As a result, increased intracellular production of ROS and excessive cell death were recorded. The so observed changes were suppressed by the administration of a ferroptosis inhibitor -

ferrostatin-1. In the same study, in cardiomyocytes with excessive iron accumulation, no molecular changes typical of autophagy or apoptosis were detected [34].

Authors also [34] report the protective role of cardiomyocytes of the mammalian target of rapamycin (mTOR) - a key effector in the insulin signaling pathway, which regulates not only cellular metabolism but also cell survival against excess iron and ferroptosis.

In another study was reported cell death with the morphological features of ferroptosis [32]. The experiment was conducted in rats with heart failure that was experimentally induced by ligation of the thoracic aorta and in the H9c2 myocytes (a subclone of an original clonal cell line, derived from embryonic rat heart tissue) in vitro, treated with erastin or isoproterenol (ISO). The described cell death is characterized by reduced vitality of the cell, increased lipid peroxidation and unstable iron deposits. Both rats and H9c2 cells were treated with puerarin - a phytoestrogen with antioxidant properties, approved for the treatment of cardiovascular and other diseases in China [35]. Interestingly, lipid peroxidation and iron overload are significantly blocked, following the administration of puerarin (phytoestrogen with antioxidant properties) [32]. These results provide convincing evidence that puerarin reduces myocyte loss during HF, partly by reducing ferroptosis, suggesting a new mechanism for potential therapy for heart failure.

## CONCLUSION

Despite the available studies in this area, there is now no definitive evidence of the involvement of iron-mediated ferroptosis in the pathogenesis of various cardiovascular diseases. The confirmation of this role is likely to encourage and improve the search for new therapeutic options.

---

## REFERENCES:

1. Ray PD, Huang BW, Tsuji Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal*. 2012 May;24(5):981-90. [[PubMed](#)] [[Crossref](#)]
2. Dixon SJ, Lemberg KM, Lamprecht MR, Skouta R, Zaitsev EM, Gleason CE, et al. Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell*. 2012 May 25;149(5):1060-72. [[PubMed](#)] [[Crossref](#)]
3. Yang WS., SriRamaratnam R., Welsch ME., Shimada K., Skouta R., Viswanathan VS, et al. Regulation of ferroptotic cancer cell death by GPX4. *Cell*. 2014 Jan; 156(1-2): 317-31. [[PubMed](#)] [[Crossref](#)]
4. Lachaier E, Louandre C., Godin C., Saidak Z, Baert M, Diouf M, et al. Sorafenib induces ferroptosis in human cancer cell lines originating from different solid tumors. *Anticancer Res*. 2014 Nov;34(11): 6417-22. [[PubMed](#)]
5. Ooko E., Saeed ME, Kadioglu O, Sarvi S, Colak M, Elmasaoudi K, et al. Artemisinin derivatives induce iron-dependent cell death (ferroptosis) in tumor cells. *Phytomedicine*. 2015 Oct; 22(11): 1045-54. [[PubMed](#)] [[Crossref](#)]
6. Yang WS, Stockwell BR. Ferroptosis: Death by Lipid Peroxidation. *Trends Cell Biol*. 2016 Mar; 26(3): 165-76. [[PubMed](#)] [[Crossref](#)]
7. Eling N, Reuter L, Hazin J, Hamacher-Brady A, Brady NR. Identification of artesunate as a specific activator of ferroptosis in pancreatic cancer cells. *Oncoscience* 2015. May; 2(5): 517-32. [[PubMed](#)] [[Crossref](#)]
8. Louandre C, Marcq I, Bouhlal H, Lachaier E., Godin C, Saidak Z, et al. The retinoblastoma (Rb) protein regulates ferroptosis induced by

- sorafenib in human hepatocellular carcinoma cells. *Cancer Lett.* 2015 Jan; 356(2 Pt B): 971-7. [[PubMed](#)] [[Crossref](#)]
9. Dolma S, Lessnick SL, Hahn WC, Stockwell BR. Identification of genotype-selective antitumor agents using synthetic lethal chemical screening in engineered human tumor cells. *Cancer cell.* 2003 Mar; 3(3): 285-96. [[PubMed](#)]
  10. Yang WS, Stockwell BR. Synthetic lethal screening identifies compounds activating iron-dependent, nonapoptotic cell death in oncogenic-RAS-harboring cancer cells. *Chem Biol.* 2008 Mar; 15(3): 234-45. [[PubMed](#)] [[Crossref](#)]
  11. Louandre C, Ezzoukhry Z, Godin C, Barbare JC, Maziere JC, Chauffert B, et al. Iron-dependent cell death of hepatocellular carcinoma cells exposed to sorafenib *Int J Cancer.* 2013 Oct; 133(7): 1732-42. [[PubMed](#)] [[Crossref](#)]
  12. Abrams RP, Carroll WL, Woerpel KA. Five-Membered Ring Peroxide Selectively Initiates Ferroptosis in Cancer Cells. *ACS Chem Biol.* 2016 May; 11(5): 1305-12. [[PubMed](#)] [[Crossref](#)]
  13. Cao JY, Dixon SJ. Mechanisms of ferroptosis. *Cell Mol Life Sci.* 2016 Jun; 73(11-12): 2195-209. [[PubMed](#)] [[Crossref](#)]
  14. Xie Y, Hou W, Song X, Yu Y, Huang J, Sun X, et al. Ferroptosis: process and function. *Cell Death Differ.* 2016 Mar; 23(3): 369-79. [[PubMed](#)] [[Crossref](#)]
  15. Kerr JF, Wyllie AH, Currie AR. Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer.* 1972 Aug; 26(4): 239-57. [[PubMed](#)]
  16. Schweichel JU, Merker HJ. The morphology of various types of cell death in prenatal tissues. *Teratology.* 1973 Jun; 7(3): 253-66. [[PubMed](#)] [[Crossref](#)]
  17. Clarke PG. Developmental cell death: morphological diversity and multiple mechanisms. *Anat Embryol (Berl).* 1990 Mar; 181(3): 195-213. [[PubMed](#)]
  18. Yu H, Guo P, Xie X, Wang Y, Chen G. Ferroptosis, a new form of cell death, and its relationships with tumorous diseases. *J Cell Mol Med.* 2017 Apr; 21(4): 648-57. [[PubMed](#)] [[Crossref](#)]
  19. Torii S, Shintoku R, Kubota C, Yaegashi M, Torii R, Sasaki M, et al. An essential role for functional lysosomes in ferroptosis of cancer cells. *Biochem J.* 2016 Mar; 473(6): 769-77. [[PubMed](#)] [[Crossref](#)]
  20. Sato H, Tamba M, Ishii T, Bannai S. Cloning and expression of a plasma membrane cystine/glutamate exchange transporter composed of two distinct proteins. *J Biol Chem.* 1999 Apr; 274(17): 11455-8. [[PubMed](#)]
  21. Bridges RJ, Natale NR, Patel SA. System xc(-) cystine/glutamate antiporter: an update on molecular pharmacology and roles within the CNS. *Br J Pharmacol.* 2012 Jan; 165(1): 20-34. [[PubMed](#)] [[Crossref](#)]
  22. Lo M, Wang YZ, Gout PW. The xc(-) cystine/glutamate antiporter: a potential target for therapy of cancer and other diseases *J Cell Physiol.* 2008 Jun; 215(3): 593-602. [[PubMed](#)] [[Crossref](#)]
  23. Tan S, Schubert D, Maher P. Oxytosis: A novel form of programmed cell death. *Curr Top Med Chem.* 2001 Dec; 1(6): 497-506. [[PubMed](#)]
  24. Dixon SJ, Stockwell BR. The role of iron and reactive oxygen species in cell death. *Nat Chem Biol.* 2014 Jan; 10(1): 9-17. [[PubMed](#)] [[Crossref](#)]
  25. Bannai S, Kitamura E. Transport interaction of L-cystine and L-glutamate in human diploid fibroblasts in culture. *J Biol Chem.* 1980 Mar; 255(6): 2372-6. [[PubMed](#)]
  26. Wolpaw AJ., Shimada K, Skouta R, Welsch ME, Akavia UD, Pe'er D, et al. Modulatory profiling identifies mechanisms of small molecule-induced cell death. *Proc Natl Acad Sci U S A.* 2011 Sep; 108(39): E771-80. [[PubMed](#)] [[Crossref](#)]
  27. Skouta R, Dixon SJ, Wang J, Dunn DE, Orman M, Shimada K, et al. Ferrostatins inhibit oxidative lipid damage and cell death in diverse disease models. *J Am Chem Soc.* 2014 Mar; 136(12): 4551-6. [[PubMed](#)] [[Crossref](#)]
  28. Kremastinos DT, Farmakis D. Iron overload cardiomyopathy in clinical practice. *Circulation.* 2011 Nov; 124(20): 2253-63. [[PubMed](#)] [[Crossref](#)]
  29. Miranda CJ, Makui H, Soares RJ, Bilodeau M, Mui J, Vali H, et al. Hfe deficiency increases susceptibility to cardiotoxicity and exacerbates changes in iron metabolism induced by doxorubicin. *Blood* 2003. Oct; 102(7): 2574-80. [[PubMed](#)] [[Crossref](#)]
  30. Nitenberg A, Ledoux S, Valensi P, Sachs R, Antony I. Coronary microvascular adaptation to myocardial metabolic demand can be restored by inhibition of iron-catalyzed formation of oxygen free radicals in type 2 diabetic patients. *Diabetes* 2002. Mar; 51(3): 813-8. [[PubMed](#)]
  31. Lapenna D, Ciofani G, Pierdomenico SD, Giamberardino MA, Porreca E. Iron status and oxidative stress in the aged rabbit heart. *J Mol Cell Cardiol.* 2018 Jan; 114: 328-33. [[PubMed](#)] [[Crossref](#)]
  32. Liu B, Zhao C, Li H, Chen X, Ding Y, Xu S. Puerarin protects against heart failure induced by pressure overload through mitigation of ferroptosis. *Biochem Biophys Res Commun.* 2018 Feb; 497(1): 233-40. [[PubMed](#)] [[Crossref](#)]
  33. Gao M, Monian P, Quadri N, Ramasamy R, Jiang X. Glutaminolysis and Transferrin Regulate Ferroptosis. *Mol Cell.* 2015 Jul; 59(2): 298-308. [[PubMed](#)] [[Crossref](#)]
  34. Baba Y, Higa JK, Shimada BK, Horiuchi KM, Suhara T, Kobayashi M, et al. Protective effects of the mechanistic target of rapamycin against excess iron and ferroptosis in cardiomyocytes. *Am J Physiol Heart Circ Physiol.* 2018 Mar 1; 314(3): H659-H668. [[PubMed](#)] [[Crossref](#)]
  35. Zhou YX, Zhang H, Peng C. Puerarin: a review of pharmacological effects. *Phytother Res.* 2014 Jul; 28(7): 961-75. [[PubMed](#)] [[Crossref](#)].

*Please cite this article as:* Kashlov Y, Dimova M. Ferroptosis and its potential role in Cardiovascular diseases. *J of IMAB*. 2019 Jan-Mar;25(1):2414-2418. DOI: <https://doi.org/10.5272/jimab.2019251.2414>

Received: 24/12/2018; Published online: 14/03/2019



**Address for correspondence:**

Yavor Kashlov,  
First Clinic of Cardiology, Department of Internal Medicine, University Hospital St. Marina, Varna.  
1, Hr. Smirnensky blvd., Varna, Bulgaria.  
Tel.: +359896084083  
E-mail: [javork@abv.bg](mailto:javork@abv.bg)